One • Step®((IVD

DUS 8 / DUS 10 / DUS 11 REAGENT STRIPS FOR URINALYSIS

PRODUCT NAME: Urine Reagent Strip

The strips you have will test either 8, 10 or 11 different parameters in urine as detailed below. Please check the label on the test tub to see which test you have (DUS 8, DUS 10 or DUS 11).

ONE+STEP® DUS 8

Reagent Strips for the rapid determination of Leukocytes. Nitrite. Protein, pH, Blood, S.G, Ketone, Glucose

ONF+STFP® DUS 10

Reagent Strips for the rapid determination of Leukocytes. Nitrite. Urobilinogen, Protein, pH, Blood, S.G, Ketone, Bilirubin, Glucose

• ONE+STEP® DUS 11

Reagent Strips for the rapid determination of Leukocytes. Nitrite. Urobilinogen, Protein, pH, Blood, S.G. Ketone, Bilirubin, Glucose, Ascorbic Acid

SUMMARY AND EXPLANATION

ONE+STEP® DUS Reagent Strips are dip-and-read test strips for In Vitro Diagnostic Use only for testing the above items in urine. Test result may provide information regarding the status of carbohydrate metabolism, kidney and liver function, acid-base balance, and urinary tract infection. It is measured by comparison of test paper attached to a plastic strip with the colour chart blocks printed on the vial label. The strips may be read visually. They can also be read instrumentally, using urine chemistry analyzers.

WARNING AND PRECAUTIONS

For in vitro diagnostic use only.

For professional use only.

CHEMICAL PRINCIPLES OF PROCEDURE AND INGREDIENTS

Urobilinogen (DUS 10 & DUS 11 only): The test is based on the Ehrlich's reaction

Ingredients: 4-Methoxybenzenediazonium 2.9mg

Glucose: Glucose oxidase catalyzes the oxidation of glucose to form hydrogen peroxide. The hydrogen peroxide thus formed then oxidizes a chromogen on the reaction pad by the action of peroxidase.

Ingredients: Glucose oxidase 430U. Peroxidase 200U. Potassium

Bilirubin (DUS 10 & DUS 11 only): Azo-coupling reaction of bilirubin with a diazonium salt in an acid medium to form an azodye.

Ingredients: Sodium nitrite 0.733 mg, 2.4-dichlorobenzene diazonium 2.3mg. Sulfosalicylic acid 25mg

Ketones: Legal's test-nitroprusside reaction. Acetoacetic acid in an alkaline medium reacts with nitroferricanide

Ingredients: Sodium nitroprusside 23.0mg

pH: This test is based on a double indicator principle that gives a broad range of colours covering the entire urinary pH range, (pH 5.0 to 9.0) Ingredients: Methyl red 0.05mg, Bromothymol blue 0.5mg

Blood: The peroxidase-like action of hemoglobin and myoglobin

specifically catalyzes the oxidation of the indicator by means of the organic hydroperoxide contained in the test paper to give a blue colouration.

Ingredients: Cumene Hydroperoxide 12mg, o-Tolidine 35mg

Specific Gravity (SG): Ionic solutes present in the urine cause protons to be released from a polyelectrolyte. As the protons are released the pH decreases and produces a colour change of bromothymol blue from blue-green to vellow-green.

Ingredients: Bromothymol blue 0.5mg

Poly vinyl ether-ALT-maleic acid anhydrous 140.5mg

Protein: This test is based on the principle of the protein error of pH indicators. At a constant pH, the development of any green colour is due to the presence of protein.

Ingredients: Tetrabromophenol blue 0.34mg

Nitrite: The test is based on the diazotization reaction of nitrite with an aromatic amine to produce a diazonium salt. It is followed by an azocoupling reaction of this diazonium salt with an aromatic compound on the reaction pad. The azo dve produced causes a pink colour change.

Ingredients: P-arsanilic acid 4.5mg

Leukocyte: This test reveals the presence of granulocyte esterases. These esterases cleave an indoxyl ester, and the indoxyl so liberated reacts with a diazonium salt to produce a violet dve.

Ingredients: Induced Indole amino acid ester 1.3mg

Ascorbic acid (DUS 11 only): The test field involves the decolourization of Tillmann's reagent.

Ingredients: 2.6-dichloro indophenol sodium salt 0.8mg

STORAGE AND HANDLING

Store in a cool, dry place at temperatures between 2°C ~ 30°C. Do not store the strips in a refrigerator or freezer. Store away from moisture and light. When stored in the original container, the product is stable up to the expiry date printed on the label and (or) vial box. Replace the bottle cap immediately and tightly after removing test strips, and keep the vial tightly closed between tests. Do not remove desiccant from bottle. Do not touch test areas of urine reagent strips. Do not open

container until ready to use.

Discolouration or darkening of the test pads may indicate deterioration. If this is evident, or if test results are questionable or inconsistent with expected finding, confirm that the product is within its expiration date and is reacting properly using known negative and positive control materials. Do not use after the expiry date. Please note that once the canister has been opened, the remaining strips remain stable for up to 6

SPECIMEN COLLECTION AND PREPARATION

Collect urine in a clean, dry container that allows complete immersion of all the fields on the test strip. Do not add preservatives. Test the specimen as soon as possible, with the sample well mixed but not centrifuged. The use of fresh morning urine is recommended for optimal nitrite tests, as well as for the valid determination of bilirubin and urobilinogen, since these compounds are unstable when exposed to light. If immediate testing is not possible, the sample should be stored in the refrigerator, but not frozen, and then brought to room temperature before used in the test. Unpreserved urine at room temperature may undergo pH changes due to microbial proliferation, which may interfere with protein determination. If cleanly voided specimens are not collected from females, positive results for leukocytes may be found due to contamination from outside the urinary tract. Skin cleansers containing chlorhexidine may affect protein test results if specimen contamination occurs.

VISUAL TEST PROCEDURE

The procedure must be followed exactly to achieve reliable results. Do not compare strips with colour chart before the strip is dipped in urine.

- 1) Dip the strip into the urine up to the test area for no more than two
- 2) Draw the edge of the strip along the brim of the vessel to remove excess urine, but make sure the reagent pads do not come into contact with the brim of the vessel.

Turn the strip on its side and tap once on a piece of absorbent material to remove any remaining urine: Excessive urine on the strip may cause the interaction of chemicals between adjacent reagent pads, so that an incorrect result may occur.

3) Compare the colours of the reagent pads exactly after 60 seconds (Leukocytes after 90~120 seconds) with the colour chart on the vial label under good light. While comparing, keep the strip horizontally to prevent possible mixing of chemicals when excessive urine is present.



For best results, performance of reagent strips should be confirmed by testing known negative and positive specimen or controls (e.g., Quantimetrix Dipper Urine Dipstick, Dropper Urine Dipstick, Dip&Spin Urine Dipstick; Bio-Rad qUAntify Plus Control; Thermo SCIENTIFIC MAS UA Control) whenever a new bottle is first opened. Each laboratory should establish its own goals for adequate standards of performance. Each lab worker should ensure that it complies with government and local requirements.

LIMITATIONS OF PROCEDURE

As with all laboratory tests, definitive diagnostic or therapeutic decisions should not be based on any single result of method. Substances that cause abnormal urine colour may affect the readability of test pads in urinalysis reacent strips.

Urobilinogen (DUS 10 & DUS 11 only): The absence of urobilinogen in the specimen cannot be determined. The test area will react with interfering substances known to react with Ehrlich's reagent, such as paminosalicylic acid. Drugs containing azo gantrisin may give a masking golden colour. The test is not reliable method for the detection of porphobilinogen.

Glucose: High SG (>1.020) with high pH urine and ascorbic acid (more than 40mg/dl) may cause a false negative for specimen containing small amounts of glucose (100mg/dl). Reactivity may be influenced by urine SG and temperature.

Billirubin (DUS 10 & DUS 11 only): Metabolities of drugs, such as pyridum and selenium, which give a colour at low pH, may cause false positives. Indican (indoxyl sulfate) can produce a yellow-orange to red colour response, which may interfere with the interpretation of negative or positive bilirubin readings. Ascorbic acid (> 30mg/dl) may cause false negative result.

Ketones: Positive results (trace or less) may occur with highly pigmented urine specimens or those containing large amounts of levodopa metabolites. Some high SG and low pH urine may give false positive result. Phenosulfonphthalein may cause false positive result.

pH: If the excessive urine remains on the strip because of improper test procedure, it is possible that the acidic buffer in protein portion comes out and affects the pH portion, so pH result may be more than the actual. This phenomenon is called "run-over effect."

Blood: Elevated specific gravity or protein in urine may reduce the reactivity of the blood test portion. Microbial peroxidase associated with urinary tract infection may cause false positive results. Ascorbic acid concentrations (>30 mg/dl) may cause false negatives at the low level of blood.

Specific Gravity (SG): High-buffered alkaline urine may cause diminished result, whereas high-buffered acidic urine may cause slightly elevated result.

Protein: False positive results may be found in strongly basic urine (pH 9). The interpretation of results is also difficult in turbid urine specimens. Nitrite: Ascorbic acid (>30mg/dL) may cause false negative result with ow level of nitrite containing (<0.03mg) urine. The negative result does not always mean that the patient is free from bacteriuria. Pink soots or

pink edges should not be interpreted as a positive result. Negative result may occur when urinary tract infections are caused by organism which do not contain nitrate reductase, when urine has not been retained in the bladder long enough (four hours or more) for reduction of nitrate to nitrite occur, or when dietary nitrate is absent.

Leukocyte: The test result may not always be consistent with the leukocyte cell number by the microscopic examination. High concentration of glucose, high specific gravity, high level of albumin, high concentration of formaldehyde or presence of blood may cause decreased test results. False positive results may occasionally be due to contamination of the specimen by vacinal discharge.

Ascorbic acid (DUS 11 only): No interferences are known.

EXPECTED VALUES

Urobilinogen (DUS 10 & DUS 11 only): The normal urobilinogen range is 0.1 to 1.0 Ehrlich unit /dl. If results exceed the concentration of 2.0 mg/dl, the patient and the urine specimen should be evaluated further.

Glucose: The kidney normally excretes small amounts of glucose. Concentrations of 100mg/dl may be considered as abnormal if found consistently.

Bilirubin (DUS 10 & DUS 11 only): Normally no bilirubin is detectable in urine by even the most sensitive methods. Even trace amounts of bilirubin are sufficiently abnormal to require further investigation.

Ketones: Ketone bodies should not be detected in normal urine specimens with this reagent.

pH: Urine values generally range from pH 5 to 9.

Blood: Normally, no haemoglobin is detectable in urine (0.010mg/dl; 3 RBC/µl). When haemoglobin appears in urine it indicates kidney disease or a urinary tract disorder. Blood may often be found in the urine of menstruating females.

Specific Gravity (SG): The normal SG of urine ranges from 1.001 to 1.035.

Protein: Normal urine specimens ordinarily contain some protein (<20mg/dL) therefore only persistent elevated levels of urine protein indicate kidney or urinary tract disease. The persistent results of trace level or over indicate significance proteinuria and thus further clinical testing is needed to evaluate the significant of results.

Nitrite: Normally no nitrite is detectable in urine.

Leukocyte: Normally no leukocytes are detectable in urine.

Ascorbic acid (DUS 11 only): The average daily intake ranges from 30-80mg, with an output of 20-30mg/day.

PERFORMANCE CHARACTERISTICS

Performance characteristics are based on clinical and analytical studies and depend upon several factors: the variability of colour perception; the presence or absence of inhibitory and matrix factors typically found in urine; and the laboratory conditions in which the product is used (e.g., lighting, temperature and humidity). Each colour block represents a range of values. Because of specimen and reading variability, specimens with analyte concentrations that fall between normal levels may give results at either level. Results will usually be within one level of the true concentration. The following list shows the generally

detectable levels of the analytes in contrived urines; however, because of the inherent variability of clinical urines, lesser concentrations may be detected under certain conditions.

TEST PAD AND SENSITIVITY (SPECIFICITY)

 Glucose:
 75-125mg/dL (Glucose)

 Bilirubin:
 0.8-1.0mg/dL (Bilirubin)

 Ketones:
 5-10mg/dL (Acetoacetic acid)

 Blood:
 10-15 RBC/µl (hemoglobin)

 Protein:
 15-30mg/dL (albumin)

 Nitrite:
 0.05-0.1mg/dL (Nitrite ion)

Leukocytes: 20-25 WBC/ul (Intact and lysed WBCs)

BIBI IOGRAPHY

- NCCLS (National Committee for Clinical Laboratory Standard) GP 16-A/ ROUTINE URINALYSIS AND COLLECTION TRANSPORATION AND PRESEVATION OF URINE SPECIMENS; TRNTATIVE GUIDELINE VOL 12-NO 26. EC.1992

NOTES ON SYMBOLS

Consult instructions for use

In vitro diagnostic

Use By (Expiry Date(YYYY-MM))

Do not reuse

Store at

Keep away from sunlight

V Number of test strips

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